

Increasing crop heterogeneity enhances multitrophic diversity across agricultural regions

Clélia Sirami^{a,b,c,1}, Nicolas Gross^{d,e,f}, Alette Bøsem Baillod^{g,h,2}, Colette Bertrand^{i,j,k,2}, Romain Carrié^{b,l,2}, Annika Hass^{g,2}, Laura Henckel^{e,m,n,o,2}, Paul Miguet^{e,m,n,p,2}, Carole Vuillot^{a,q,2}, Audrey Alignier^{i,r}, Jude Girard^s, Péter Batáry^{g,t}, Yann Clough^{g,l}, Cyrille Violle^a, David Giralt^u, Gerard Bota^u, Isabelle Badenhauer^{e,m,n,v}, Gaëtan Lefebvre^w, Bertrand Gauffre^{e,m,n}, Aude Vialatte^{b,c}, François Calatayud^{b,c}, Assu Gil-Tena^x, Lutz Tischendorf^s, Scott Mitchell^s, Kathryn Lindsay^s, Romain Georges^k, Samuel Hilaire^w, Jordi Recasens^{y,z}, Xavier Oriol Solé-Senar^{y,z}, Irene Robledo^{y,z}, Jordi Bosch^{aa}, Jose Antonio Barrientos^{bb}, Antonio Ricarte^{cc}, Maria Angeles Marcos-Garcia^{cc}, Jesús Miñano^{dd}, Raphaël Mathevet^a, Annick Gibon^b, Jacques Baudry^{i,r}, Gérard Balent^b, Brigitte Poulin^w, Françoise Burel^{kr}, Teja Tschamtker^g, Vincent Bretagnolle^{e,m}, Gavin Siriwardena^{ee}, Annie Ouin^{b,c}, Lluís Brotons^{u,x,ff}, Jean-Louis Martin^{a,3}, and Lenore Fahrig^{s,3}

^aUMR 5175 CEFE, CNRS, Université de Montpellier, Université Paul Valéry Montpellier, EPHE, IRD, F-34293 Montpellier, France; ^bUMR 1201 DYNAFOR, Université de Toulouse, INRA, F-31326 Castanet-Tolosan, France; ^cLTSE Zone Atelier Pyrénées Garonne, F-31320 Auzeville-Tolosane, France; ^dUMR Ecosystème Prairial, UCA, INRA, VetAgro Sup, F-63000 Clermont-Ferrand, France; ^eLTSE Zone Atelier Plaine et Val de Sèvre, F-79360 Villiers en Bois, France; ^fDepartamento de Biología y Geología, Física y Química Inorgánica, Escuela Superior de Ciencias Experimentales y Tecnología, Universidad Rey Juan Carlos, 28933 Móstoles, Spain; ^gAgroecology, Department for Crop Sciences, University of Göttingen, D-37077 Göttingen, Germany; ^hAgroecology and Biodiversity, Research Institute of Organic Agriculture, CH-5070 Frick, Switzerland; ⁱUMR 0980 BAGAP, INRA, Agrocampus Ouest, ESA, F-35042 Rennes, France; ^jUMR 1402 ECOSYS, INRA, AgroParisTech, Université Paris-Saclay, F-78026 Versailles, France; ^kUMR 6553 ECOBIO, CNRS, Université de Rennes 1, F-35042 Rennes, France; ^lCentre for Environmental and Climate Research, Lund University, SE-22362 Lund, Sweden; ^mUMR 7372 CEBEC, CNRS, Université de La Rochelle, F-79360 Villiers en Bois, France; ⁿUSC1339 SEC-LR, INRA, Villiers en Bois, F-79360 Niort, France; ^oSwedish Species Information Centre, Swedish University of Agricultural Sciences, SE-75007 Uppsala, Sweden; ^pUR1115 PSH, INRA, F-84914 Avignon, France; ^qUMR 7204 CESCO, Sorbonne Universités, CNRS, UPMC, Muséum National d'Histoire Naturelle, F-75005 Paris, France; ^rLTSE, Zone Atelier Armorique, F-35042 Rennes Cedex, France; ^sGeomatics and Landscape Ecology Laboratory, Carleton University, Ottawa, ON K1S 5B6, Canada; ^t"Lendület" Landscape and Conservation Ecology, Institute of Ecology and Botany, MTA Centre for Ecological Research, 2163 Vácrátót, Hungary; ^uCTFC Forest Sciences Centre of Catalonia, 25280 Solsona, Spain; ^vURP3F, INRA, 86600 Lusignan, France; ^wTour du Valat, Research Institute for the Conservation of Mediterranean Wetlands, F-13200 Arles, France; ^xInForest Joint Research Unit, 25280 Solsona, Spain; ^yAgrotecnio, Universitat de Lleida, 25198 Lleida, Spain; ^zDepartment of HBJ, ETSEA, Universitat de Lleida, 25198 Lleida, Spain; ^{aa}CREAF, 08193 Cerdanyola del Vallès, Spain; ^{bb}Departament de Biologia Animal, de Biologia Vegetal i d'Ecologia, Universitat Autònoma Barcelona, 08193 Cerdanyola del Vallès, Spain; ^{cc}Instituto Universitario de Investigación, CIBIO, University of Alicante, 03690 San Vicente del Raspeig, Spain; ^{dd}Area de Ecología, Facultad de Biología, Universidad de Murcia, 30100, Murcia, Spain; ^{ee}British Trust for Ornithology, Thetford, Norfolk IP24 2PU, United Kingdom; and ^{ff}CSIC, 08193 Cerdanyola del Vallès, Spain

Edited by Claire Kremen, University of California, Berkeley, CA, and accepted by Editorial Board Member Ruth S. DeFries June 28, 2019 (received for review April 18, 2019)

Agricultural landscape homogenization has detrimental effects on biodiversity and key ecosystem services. Increasing agricultural landscape heterogeneity by increasing seminatural cover can help to mitigate biodiversity loss. However, the amount of seminatural cover is generally low and difficult to increase in many intensively managed agricultural landscapes. We hypothesized that increasing the heterogeneity of the crop mosaic itself (hereafter "crop heterogeneity") can also have positive effects on biodiversity. In 8 contrasting regions of Europe and North America, we selected 435 landscapes along independent gradients of crop diversity and mean field size. Within each landscape, we selected 3 sampling sites in 1, 2, or 3 crop types. We sampled 7 taxa (plants, bees, butterflies, hoverflies, carabids, spiders, and birds) and calculated a synthetic index of multitrophic diversity at the landscape level. Increasing crop heterogeneity was more beneficial for multitrophic diversity than increasing seminatural cover. For instance, the effect of decreasing mean field size from 5 to 2.8 ha was as strong as the effect of increasing seminatural cover from 0.5 to 11%. Decreasing mean field size benefited multitrophic diversity even in the absence of seminatural vegetation between fields. Increasing the number of crop types sampled had a positive effect on landscape-level multitrophic diversity. However, the effect of increasing crop diversity in the landscape surrounding fields sampled depended on the amount of seminatural cover. Our study provides large-scale, multitrophic, cross-regional evidence that increasing crop heterogeneity can be an effective way to increase biodiversity in agricultural landscapes without taking land out of agricultural production.

multitaxa | biodiversity | crop mosaic | farmland | landscape complementation

Agriculture dominates the world's terrestrial area (1). Agricultural landscape homogenization through the decrease of seminatural cover, crop specialization, and field enlargement (2–4) represents a continuing worldwide threat to biodiversity and the

delivery of key ecosystem services to people (5). There is ample evidence that enhancing landscape heterogeneity by reversing

Significance

Agricultural landscape homogenization is a major ongoing threat to biodiversity and the delivery of key ecosystem services for human well-being. It is well known that increasing the amount of seminatural cover in agricultural landscapes has a positive effect on biodiversity. However, little is known about the role of the crop mosaic itself. Crop heterogeneity in the landscape had a much stronger effect on multitrophic diversity than the amount of seminatural cover in the landscape, across 435 agricultural landscapes located in 8 European and North American regions. Increasing crop heterogeneity can be an effective way to mitigate the impacts of farming on biodiversity without taking land out of production.

Author contributions: G. Balent, B.P., F.B., T.T., V.B., G.S., A.O., L.B., J.-L.M., and L.F. designed research; C.S., N.G., A.B.B., C.B., R.C., A.H., L.H., P.M., C. Vuillot, A.A., J.G., P.B., Y.C., C. Violle, D.G., G. Bota, I.B., G.L., B.G., A.V., F.C., A.G.-T., L.T., S.M., K.L., R.G., S.H., J.R., X.O.S.-S., I.R., J. Bosch, J.A.B., A.R., M.A.M.-G., J.M., R.M., A.G., and J. Baudry performed research; C.S., N.G., A.B.B., C.B., R.C., A.H., L.H., and P.M. analyzed data; and C.S. and N.G. wrote the paper.

The authors declare no conflict of interest.

This article is a PNAS Direct Submission. C.K. is a guest editor invited by the Editorial Board.

Published under the PNAS license.

A previous version of this work was published as part of Annika Hass's 2018 Doctoral dissertation thesis, Farmland heterogeneity effects on biodiversity, community traits and insect pollination. Georg-August-Universität Göttingen, pp 16–67.

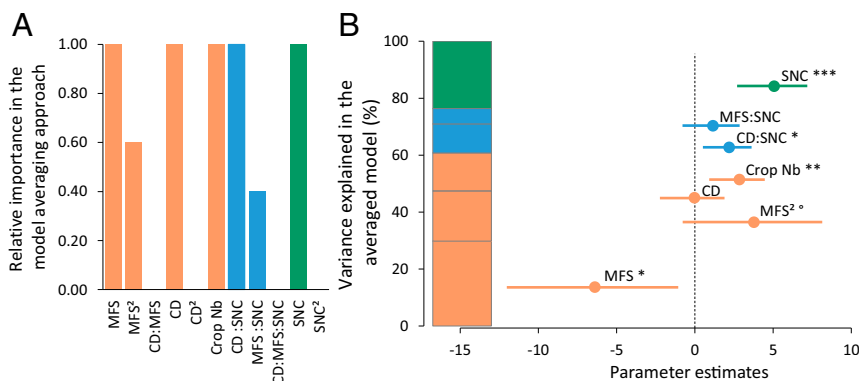
¹To whom correspondence may be addressed. Email: clelia.sirami@inra.fr.

²A.B.B., C.B., R.C., A.H., L.H., P.M., and C. Vuillot contributed equally to this work.

³J.-L.M. and L.F. contributed equally to this work.

This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1906419116/-DCSupplemental.

Published online July 29, 2019.



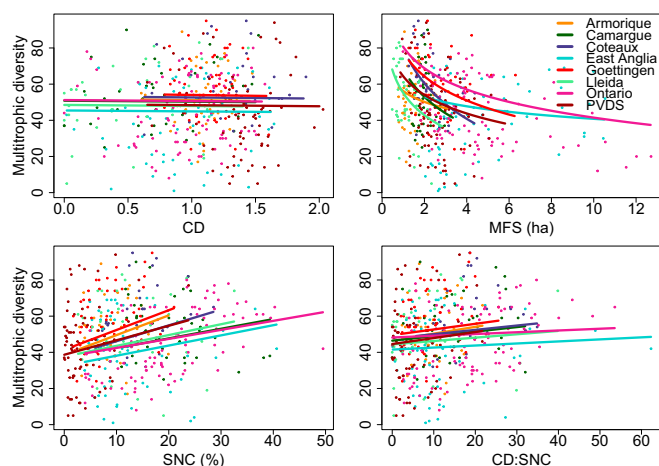


Fig. 4. Effects of CD, MFS, SNC, and the interaction between CD and SNC (CD:SNC) on multitrophic diversity in different regions. Slopes are based on the outputs of model 2 (see *Methods*) including a random effect of region on these 4 slopes ($n = 435$ landscapes). Colors indicate the region.

The positive landscape-level effect of increasing crop diversity on multitrophic diversity observed in landscapes with more than 11% seminatural cover supports the “landscape complementation” hypothesis (Hyp-1b in *SI Appendix*, section 1). This finding is consistent with the fact that a diverse crop matrix provides a temporal continuity of food sources (31) while seminatural patches provide stable resources, for example, for nesting or shelter (e.g., ref. 32). Such complementation among multiple cover types has been described for several species (e.g., refs. 33 and 34). Our study, based on multiple regions and multiple trophic groups, shows that the positive landscape-level effect of increasing crop diversity can be generalized to multitrophic diversity across many agricultural landscapes (50% of landscapes included in our study).

The negative landscape-level effect of increasing crop diversity on multitrophic diversity in landscapes with less than 4% seminatural cover supports the “minimum total habitat area requirement” hypothesis (Hyp-1c in *SI Appendix*, section 1). This finding is consistent with the fact that landscape simplification tends to filter out species with large body sizes (35), which also have high minimum total habitat area requirements (36), and may therefore require high amount of a single crop type. However, taxa included in the present study were associated with a wide range of ecological traits, and therefore a wide range of minimum total habitat area requirements. That they showed a consistent response to crop diversity and the interaction of crop diversity and seminatural cover (Fig. 5) suggests that the minimum total habitat area requirement hypothesis is unlikely to be the sole mechanism contributing to our results. Future research is needed to identify additional mechanisms and conditions under which increasing crop diversity leads to a consistent net positive effect on multitrophic diversity, i.e., a positive effect of habitat specialization plus landscape complementation processes.

Implications for Agricultural Policies. Our study has important implications for large-scale policy schemes implemented across a wide range of contexts such as the European Common Agricultural Policy and its recent greening (21), the Canadian Agriculture Policy Frameworks (37), or the United States Farm Bill (38).

First, our results suggest that increasing crop heterogeneity may have a similar or greater benefit for multitrophic diversity to increasing seminatural cover (Fig. 2B) or even decreasing field-level land-use intensity (ref. 16; *Table S5.12*, *SI Appendix*, section 5). Given current challenges to increase seminatural cover and limit chemical use in agricultural landscapes (39), policies aiming at increasing crop heterogeneity may represent an effective and complementary way to improve biodiversity conservation in

agricultural landscapes. Policy measures favoring crop heterogeneity may be more easily implemented than policies to increase seminatural cover or reduce chemical use (40). Associated with adequate economic incentives, they may also be more favorably perceived by farmers and thus lead to higher uptake than measures requiring farmers to take land out of production (39). Such measures may also contribute to the development of frameworks that reward farmers for sustainable land stewardship.

We observed a consistent effect of crop heterogeneity on species diversity across 7 taxa representing a wide range of ecological traits, functions, and trophic levels (plants, bees, butterflies, hoverflies, carabids, spiders, and birds; Fig. 5). We observed landscapes where 6 or even all 7 taxa reached the threshold of 60% of the maximum species richness observed within a given region (Fig. 4). Our study therefore suggests that policies to increase crop heterogeneity would be an effective way to increase the diversity of all components of biodiversity simultaneously and restore multitrophic biodiversity in agricultural landscapes.

Finally, our results can contribute to the development of policies adapted to different landscape contexts. For instance, our results suggest that policy measures aimed at decreasing field sizes to below 6 ha may be particularly effective to promote multitrophic diversity in agricultural landscapes, especially in landscapes where seminatural cover is below 8%. Our results also caution against a “blind” increase of crop diversity. Measures aimed at increasing crop diversity may be effective to promote multitrophic diversity in landscapes where seminatural cover exceeds 11%. However, they are more likely to be effective in promoting multitrophic diversity across all agricultural landscapes if combined with measures promoting the restoration or maintenance of seminatural cover.

Conclusion

Our study demonstrates the importance of crop heterogeneity for multitrophic diversity in agricultural landscapes: The effect of maintaining/increasing crop heterogeneity is likely to be as important as the effect of maintaining/increasing seminatural cover. This finding suggests that field enlargement and crop specialization, especially the former, have been underestimated drivers of past and ongoing biodiversity declines. More importantly, our study shows that increasing crop heterogeneity represents a major potential lever to increase synergies between food production and biodiversity conservation.

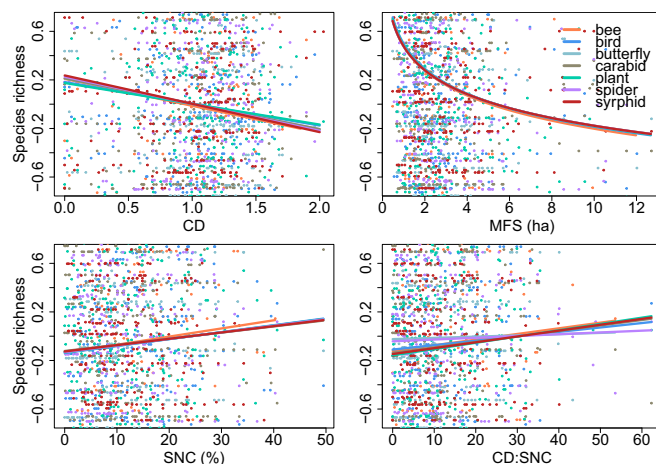


Fig. 5. Effects of CD, MFS, SNC, and the interaction between CD and SNC (CD:SNC) on the landscape-level species richness of taxonomic groups. Slopes are based on the outputs of model 10 (see *SI Appendix*, section 5) including a random effect of taxon on these 4 slopes ($n = 435$ landscapes). Colors indicate the taxon.

Methods

Region, Landscape, and Sampling Site Selection. We selected 8 agricultural regions (Armorique, Camargue, Coteaux de Gascogne, and Plaine et Val de Sèvre in France, East Anglia in the United Kingdom, Goettingen in Germany, Lleida in Spain, and Eastern Ontario in Canada; Fig. S2.1, *SI Appendix*, section 2) belonging to 6 different ecoregions (41) and differing in topography, climate, field shapes, and agricultural cover types and products (e.g., rice, dairy, tree crops).

We used the best spatial data available within each region before field work to identify all 1 km × 1-km rural landscapes, i.e., those dominated by agricultural cover (>60%, including all crops and grassland managed for agricultural production). We then developed a protocol to select a combination of landscapes that maximized the gradients of crop diversity and mean field size, while minimizing the correlation between them (42). Crop diversity may theoretically be constrained by the number and size of fields in landscapes with large fields. However, in our dataset, mean field size was smaller than 12 ha and was therefore not a limiting factor for crop diversity within the 1 km × 1-km landscapes. We selected between 32 and 93 landscapes within each region, totaling 435 landscapes across all regions.

We selected 3 sampling sites within each landscape, totaling 1,305 sampling sites across all regions. The number of crop types sampled ranged from 1 to 3 per landscape. Where feasible, we located sampling sites in dominant agricultural cover types within each region (e.g., wheat fields and oilseed rape in Goettingen). When this was not feasible, we located sampling sites in agricultural cover types that were accessible within a given landscape (*SI Appendix*, section 3). The 3 sampling sites were at least 200 m from each other, at least 50 m from the border of the landscape, and at least 50 m from patches of nonagricultural cover types such as forests and urban areas.

Multitaxa Sampling. We selected 7 taxa representing a wide range of ecological traits, functions, and trophic levels which, combined into a multidiversity index (see below), represent a proxy for multitrophic diversity: plants, bees, butterflies, hoverflies, carabids, spiders, and birds. All taxa were sampled using standardized sampling protocols across all regions, allowing us to test the consistency of effects across the 8 regions (*SI Appendix*, section 3).

At each sampling site, we selected 2 parallel 50-m “transects,” one located at the field edge and the other inside the field 25 m away from the first transect (Figs. S3.1 and S3.2, *SI Appendix*, section 3). Birds were sampled using point counts centered on the field-edge transect. Plants were surveyed along both transects. Butterflies were surveyed visually using timed walks along both transects. Bees and hoverflies were sampled using colored pan traps on poles erected at each end and in the center of all transects. Carabids and spiders were sampled using pitfall traps installed at each end of all transects. Captured arthropods were preserved in ethanol prior to identification. Multiple survey visits were conducted during the season when relevant (*SI Appendix*, section 3). Each landscape was sampled during 1 y and sampling of landscapes was distributed across 2 y within each region, between 2011 and 2014 (Table S3.1, *SI Appendix*, section 3).

We identified more than 167,000 individuals from 2,795 species (Table S3.2, *SI Appendix*, section 3). For each taxon, we calculated species richness at the landscape level, i.e., across all 3 sampling sites and across all visits when multiple survey visits were conducted. The average species richness per landscape varied greatly among taxa, from 5.4 for butterflies to 44.9 for plants. Correlations in average species richness between pairs of taxa were weak (<0.41), with an average correlation of 0.07 (Table S3.3, *SI Appendix*, section 3).

Multitrophic Diversity Index. A classical approach in the literature consists of calculating the average, standardized diversity across taxa (43). However, this approach has limitations (*SI Appendix*, section 3). Although very high/low values imply that all taxa exhibit high/low diversity, intermediate values are difficult to interpret as they may correspond to situations where 1) diversity values are intermediate for all taxa, or 2) diversity values are high for some taxa and low for others, i.e., trade-offs among taxa. To overcome this limitation, we used a threshold approach initially developed to aggregate multiple ecosystem functions (22).

For each taxon and each region, we identified the maximum species richness observed across all landscapes. We used the 95th percentile as the maximum observed species richness (hereafter “SR max”) to minimize the effect of outliers. Next, we identified which landscapes attained a given threshold (x) of SR max. We chose to use an intermediate threshold of 60% because intermediate thresholds have been shown to provide an effective measure of multitrophic diversity in agricultural landscapes (ref. 43 and *SI Appendix*, section 3). We then tallied the proportion of taxa that exceeded

the given threshold to produce a multidiversity index (T60.landscape) for each landscape, based on the following formula:

$$\text{Multidiversity}(\text{Tx. landscape}) = \frac{1}{n} \sum_{i=1}^n \text{SR}_i \text{ if } (x \times \text{SR max. region } j)],$$

where n is the number of taxa for which data were available in a given landscape (*SI Appendix*, section 3), SR_i is the number of species for taxon i , x is the minimum threshold to be reached, and $\text{SR max. region } j$ is the maximum species richness for taxon i in the region the landscape considered belonging to. This multidiversity index ranges between 0 and 1 (*SI Appendix*, section 3). For simplicity, we hereafter refer to “landscape-level multitrophic diversity” rather than T60.landscape.

Mean Field Size and Diversity of Crop Types in the Landscape. We used a standardized protocol across all regions to produce land-cover maps allowing us to compare consistency of effects across the 8 regions (*SI Appendix*, section 4). We conducted extensive ground-truthing surveys during the field seasons to map all fields, linear elements between adjacent fields, and non-agricultural covers. We built a common land-cover classification for the 8 regions (*SI Appendix*, section 4). We then used these standardized, detailed maps to calculate 4 explanatory variables for each landscape: diversity of crop types in the landscapes, mean field size, seminatural cover, and total length of seminatural linear elements between fields.

We calculated the Shannon diversity of agricultural cover types (hereafter “the diversity of crop types in the landscape,” CD) and the mean field size in hectares (MFS). Neither CD nor MFS was correlated with local land-use intensity (an index combining data on plowing, fertilizer, herbicide, and insecticide) or the overall composition of the crop mosaic across all regions (*SI Appendix*, section 5). CD and MFS were moderately correlated with the type of crops sampled in some regions and MFS was moderately correlated with the proportion of grassland in the crop mosaic, but none of these correlations affected our conclusions (*SI Appendix*, section 5). We calculated the percentage of seminatural cover types, i.e., woodland, open land, and wetland (SNC), in each landscape. We also calculated the total length of linear seminatural elements between fields, e.g., hedgerows, grassy margins (SNL; measured in meters). SNL and MFS were highly correlated in some regions (Table S5.6, *SI Appendix*, section 5). As a result, we did not include SNL in the main analyses and only tested the relative effect of MFS and SNL using a subset of our dataset for which MFS and SNL were not strongly correlated (*SI Appendix*, section 5).

Data Analysis. We first tested the effect of crop heterogeneity on multitrophic diversity (model 1). We fitted a linear mixed model with restricted maximum likelihood using the landscape-level multidiversity index (T60.landscape) as the response variable. We included the number of crop types sampled per landscape (CropNb), CD, MFS, and SNC as explanatory variables (see alternative hypotheses in *SI Appendix*, section 1). We included both interaction effects and quadratic effects. Due to a positive skew in the distribution of MFS, we used log MFS in all analyses. To reflect the large-scale spatial and temporal structure of our dataset, we added sampling year (Year), nested within study region (Region), as a random effect. To reflect the spatial structure of our dataset within each region, we included the longitude and latitude of the center of each landscape (Lat, Lon) as covariates. We standardized all fixed effects to allow for a direct comparison of estimates.

Model 1: $\text{Imer}(\text{T60.landscape} \sim \text{CD} * \text{MFS} * \text{SNC} + \text{CD}^2 + \text{MFS}^2 + \text{SNC}^2 + \text{CropNb} + \text{Lat} + \text{Lon} + (1|\text{Region}/\text{Year}))$. Then, we added random effects for region on the slopes of the diversity of crop types in the landscape, MFS, SNC, as well as the interaction between the CD and SNC (model 2). We assumed that the effects of region on the intercept and slopes were uncorrelated. To test whether region had a significant effect on the slope of either the CD, MFS, or SNC, as well as the interaction between crop diversity and SNC, we used the function exactRLRT from package RLRsim.

Model 2: $\text{Imer}(\text{T60.landscape} \sim \text{CD} * \text{MFS} * \text{SNC} + \text{CD}^2 + \text{MFS}^2 + \text{SNC}^2 + \text{CropNb} + \text{Lat} + \text{Lon} + (1|\text{Region}/\text{Year}) + (0+\text{CD}|\text{Region})) + (0+\text{MFS}|\text{Region}) + (0+\text{SNC}|\text{Region}) + (0+\text{CD:SNC}|\text{Region}))$. We then tested the effects of crop heterogeneity on the SR of taxonomic groups (model 3). To do this, we fitted a similar model, using the landscape-level SR of taxonomic groups standardized within each taxon and region as the response variable. To reflect that species pools vary between taxa, we added taxon as a random effect.

Model 3: $\text{Imer}(\text{SR} \sim \text{CD} * \text{MFS} * \text{SNC} + \text{CD}^2 + \text{MFS}^2 + \text{SNC}^2 + \text{CropNb} + \text{Lat} + \text{Lon} + (1|\text{Region}/\text{Year}) + (1|\text{Taxon}))$. Then we added random effects for taxon on the slopes of CD, MFS, SNC, as well as the interaction between CD and SNC (model 4). We assumed that the effects of taxon on the intercept and slopes were uncorrelated. To test whether taxon had a significant effect on the slope of either

CD, MFS, SNC or the interaction between CD and SNC, we used the function exactRLRT from package RLRsim.

Model 4: $lmer(SR \sim CD * MFS * SNC + CD^2 + MFS^2 + SNC^2 + CropNb + Lat + Lon + (1|Taxon) + (1|Region/Year) + (0+CD|Taxon)) + (0+MFS|Taxon) + (0+SNC|Taxon) + (0+CD:SNC|Taxon))$. We fitted all models with the R lme4 package using LMER (44), we removed outliers using function romr.fnc from package LMERConvenienceFunctions (45), and we ran diagnostic tools to verify that residuals were independently and normally distributed, and showed no spatial autocorrelation. For each model, a multimodel inference procedure was applied using the R MuMIn package (46). This method allowed us to perform model selection by creating a set of models with all possible combinations of the initial variables and sorting them according to the Akaike Information Criterion (AIC) fitted with Maximum Likelihood (47). We selected all models with $\Delta AIC < 2$ and used the model averaging approach using LMER to estimate parameters and associated *P* values, using the function model.avg. We ran all analyses using the software R 3.4.0 (48).

We ran additional analyses to check that the composition of the crop mosaic, the proportion of grassland in the crop mosaic, and the amount of seminatural vegetation occurring between fields did not affect our conclusions (SI Appendix, section 5). We also ran complementary analyses using field-level multidiversity (T60.field) as the response variable—instead of the landscape-level multidiversity index (T60.landscape)—to check that our results hold true at the field level, in particular within a subset of cereal fields, and that the type of crop sampled or the level of land-use intensity within sampled fields did not affect our conclusions (SI Appendix, section 5). Finally, we used a moving window analysis to identify potential discontinuities in multitrophic diversity response to CD and MFS along the gradient of SNC (SI Appendix, section 5).

Data Accessibility. All protocols, datasets, and R codes used in this article can be made available upon request to the corresponding author.

ACKNOWLEDGMENTS. This research was funded by the ERA-Net BiodivERsA, with the national funders French National Research Agency (ANR-11-EBID-0004), German Ministry of Research and Education, German Research Foundation and Spanish Ministry of Economy and Competitiveness, part of the 2011 BiodivERsA call for research proposals. The UK component of this research was funded by the UK Government Department of the Environment, Food and Rural Affairs (Defra), as Project WC1034. The Canadian component of this research was funded by a Natural Sciences and Engineering Research Council of Canada Strategic Project, the Canada Foundation for Innovation, Environment Canada, and Agriculture and Agri-Food Canada. N.G. was supported by the AgreenSkills+ Fellowship programme which has received funding from the EU's Seventh Framework Programme under Grant Agreement FP7-609398 (AgreenSkills+ contract). A.G.-T. (Juan de la Cierva Fellow, JCI-2012-12089) was funded by Ministerio de Economía y Competitividad (Spain). C. Violle was supported by the European Research Council Starting Grant Project "Ecophysiological and biophysical constraints on domestication of crop plants" (Grant ERC-StG-2014-639706-CONSTRAINTS). A.R.'s position at the University of Alicante is funded by the "Vicerrectorado de Investigación y Transferencia de Conocimiento." We thank the hundreds of farmers and farm owners from all 8 regions who graciously permitted us to work on their lands. In addition to the coauthors, the project involved direct assistance from more than 150 individuals for geomatics analyses, field sampling, and species identification. We thank all of them for their huge contribution to the FarmLand project and this study. We are grateful to the GLEL (Geomatics and Landscape Ecology Laboratory) Friday Discussion Group for very helpful input. Finally, we would like to thank the editor, Sarina Macfadyen, and another anonymous reviewer for their very constructive comments on an earlier version of this paper.

1. D. Tilman *et al.*, Forecasting agriculturally driven global environmental change. *Science* **292**, 281–284 (2001).
2. E. V. White, D. P. Roy, A contemporary decennial examination of changing agricultural field sizes using Landsat time series data. *Geo* **2**, 33–54 (2015).
3. C. J. Barr, M. K. Gillespie, Estimating hedgerow length and pattern characteristics in Great Britain using Countryside Survey data. *J. Environ. Manage.* **60**, 23–32 (2000).
4. J. Aguilar *et al.*, Crop species diversity changes in the United States: 1978–2012. *PLoS One* **10**, e0136580 (2015).
5. T. Newbold *et al.*, Global effects of land use on local terrestrial biodiversity. *Nature* **520**, 45–50 (2015).
6. P. Batáry, J. Fischer, A. Báldi, T. O. Crist, T. Tschamntke, Does habitat heterogeneity increase farmland biodiversity? *Front. Ecol. Environ.* **9**, 152–153 (2011).
7. T. Miyashita, Y. Chishiki, S. R. Takagi, Landscape heterogeneity at multiple spatial scales enhances spider species richness in an agricultural landscape. *Popul. Ecol.* **54**, 573–581 (2012).
8. D. Perović *et al.*, Configurational landscape heterogeneity shapes functional community composition of grassland butterflies. *J. Appl. Ecol.* **52**, 505–513 (2015).
9. P. Batáry, L. V. Dicks, D. Kleijn, W. J. Sutherland, The role of agri-environment schemes in conservation and environmental management. *Conserv. Biol.* **29**, 1006–1016 (2015).
10. G. Lüscher *et al.*, Farmland biodiversity and agricultural management on 237 farms in 13 European and two African regions. *Ecology* **97**, 1625 (2016).
11. A. Holzschuh, C. F. Dormann, T. Tschamntke, I. Steffan-Dewenter, Mass-flowering crops enhance wild bee abundance. *Oecologia* **172**, 477–484 (2013).
12. L. Raymond *et al.*, Immature hoverflies overwinter in cultivated fields and may significantly control aphid populations in autumn. *Agric. Ecosyst. Environ.* **185**, 99–105 (2014).
13. L. Fahrig *et al.*, Functional landscape heterogeneity and animal biodiversity in agricultural landscapes. *Ecol. Lett.* **14**, 101–112 (2011).
14. A.-C. Weibull, Ö. Östman, Å. Granqvist, Species richness in agroecosystems: The effect of landscape, habitat and farm management. *Biodivers. Conserv.* **12**, 1335–1355 (2003).
15. J. B. Dunning, B. J. Danielson, H. R. Pulliam, Ecological processes that affect populations in complex landscapes. *Oikos* **65**, 169–175 (1992).
16. P. Batáry *et al.*, The former Iron Curtain still drives biodiversity-profit trade-offs in German agriculture. *Nat. Ecol. Evol.* **1**, 1279–1284 (2017).
17. T. Tschamntke *et al.*, Landscape moderation of biodiversity patterns and processes—eight hypotheses. *Biol. Rev. Camb. Philos. Soc.* **87**, 661–685 (2012).
18. F. Burel, A. Butet, Y. R. Delettre, N. M. de la Pena, Differential response of selected taxa to landscape context and agricultural intensification. *Landscape Urban Plan.* **67**, 195–204 (2004).
19. A. Ponjoan, G. Bota, S. Mañosa, Ranging behaviour of little bustard males, *Tetrax tetrax*, in the lekking grounds. *Behav. Processes* **91**, 35–40 (2012).
20. S. Gaba, B. Chauvel, F. Dessaint, V. Bretagnolle, S. Petit, Weed species richness in winter wheat increases with landscape heterogeneity. *Agric. Ecosyst. Environ.* **138**, 318–323 (2010).
21. G. Pe'er *et al.*, Agriculture policy. EU agricultural reform fails on biodiversity. *Science* **344**, 1090–1092 (2014).
22. E. Allan *et al.*, Interannual variation in land-use intensity enhances grassland multidiversity. *Proc. Natl. Acad. Sci. U.S.A.* **111**, 308–313 (2014).
23. L. Fahrig *et al.*, Farmlands with smaller crop fields have higher within-field biodiversity. *Agric. Ecosyst. Environ.* **200**, 219–234 (2015).
24. S. J. Collins, L. Fahrig, Responses of anurans to composition and configuration of agricultural landscapes. *Agric. Ecosyst. Environ.* **239**, 399–409 (2017).
25. L. Monck-Whipp, A. E. Martin, C. M. Francis, L. Fahrig, Farmland heterogeneity benefits bats in agricultural landscapes. *Agric. Ecosyst. Environ.* **253**, 131–139 (2018).
26. M. Šálek *et al.*, Bringing diversity back to agriculture: Smaller fields and non-crop elements enhance biodiversity in intensively managed arable farmlands. *Ecol. Indic.* **90**, 65–73 (2018).
27. A. L. Hass *et al.*, Landscape configurational heterogeneity by small-scale agriculture, not crop diversity, maintains pollinators and plant reproduction in western Europe. *Proc. R. Soc. B* **285**, 20172242 (2018).
28. M. Berdugo *et al.*, Aridity preferences alter the relative importance of abiotic and biotic drivers on plant species abundance in global drylands. *J. Ecol.* **107**, 190–202 (2019).
29. J. Josefsson, Å. Berg, M. Hiron, T. Pärt, S. Eggers, Sensitivity of the farmland bird community to crop diversification in Sweden: Does the CAP fit? *J. Appl. Ecol.* **54**, 518–526 (2017).
30. E. M. Olimpí, S. M. Philpott, Agroecological farming practices promote bats. *Agric. Ecosyst. Environ.* **265**, 282–291 (2018).
31. N. A. Schellhorn, V. Gagic, R. Bommarco, Time will tell: Resource continuity bolsters ecosystem services. *Trends Ecol. Evol. (Amst.)* **30**, 524–530 (2015).
32. C. Sirami, L. Brotons, J. Martin, Woodlarks Lullula arborea and landscape heterogeneity created by land abandonment. *Bird Study* **58**, 99–106 (2011).
33. S. E. Pope, L. Fahrig, N. G. Merriam, Landscape complementation and metapopulation effects on leopard frog populations. *Ecology* **81**, 2498–2508 (2000).
34. T. Mueller, N. Selva, E. Pugaczewicz, E. Prins, Scale-sensitive landscape complementation determines habitat suitability for a territorial generalist. *Ecography* **32**, 345–353 (2009).
35. S. Gámez-Virués *et al.*, Landscape simplification filters species traits and drives biotic homogenization. *Nat. Commun.* **6**, 8568 (2015).
36. M. Baguette, V. Stevens, Predicting minimum area requirements of butterflies using life-history traits. *J. Insect. Conserv.* **17**, 645–652 (2013).
37. B. J. Deaton, P. Boxall, Canadian agricultural policy in the twenty-first century: Looking back and going forward. *Can. J. Agric. Econ. Can. Dapagosagroeconomie* **65**, 519–522 (2017).
38. A. Reimer, Ecological modernization in U.S. agri-environmental programs: Trends in the 2014 farm bill. *Land Use Policy* **47**, 209–217 (2015).
39. G. Pe'er *et al.*, Adding some green to the greening: Improving the EU's ecological focus areas for biodiversity and farmers. *Conserv. Lett.* **10**, 517–530 (2017).
40. C. Rodríguez, K. Wiegand, Evaluating the trade-off between machinery efficiency and loss of biodiversity-friendly habitats in arable landscapes: The role of field size. *Agric. Ecosyst. Environ.* **129**, 361–366 (2009).
41. D. M. Olson *et al.*, Terrestrial ecoregions of the world: A new map of life on Earth a new global map of terrestrial ecoregions provides an innovative tool for conserving biodiversity. *Bioscience* **51**, 933–938 (2001).
42. J. Pasher *et al.*, Optimizing landscape selection for estimating relative effects of landscape variables on ecological responses. *Landscape Ecol.* **28**, 371–383 (2013).
43. J. E. K. Byrnes *et al.*, Investigating the relationship between biodiversity and ecosystem multifunctionality: Challenges and solutions. *Methods Ecol. Evol.* **5**, 111–124 (2014).
44. D. Bates, M. Mächler, B. M. Bolker, S. C. Walker, Fitting linear mixed-effects models using lme4. *J. Stat. Softw.* **67**, 1–48 (2015).
45. A. Tremblay, J. Ransijn, LMERConvenienceFunctions: Model selection and post-hoc analysis for (G) LMER models, R package Version 2.10 (2015). <https://CRAN.R-project.org/package=LMERConvenienceFunctions>. Accessed 14 April 2019.
46. K. Barton, MuMIn: Multi-Model Inference, R Package Version 0.12.0. [Http-Forg-Project.org](http://Forg-Project.org) (2009). <https://ci.nii.ac.jp/naid/10030574914/>. Accessed 1 August 2018.
47. A. Zuur, E. N. Ieno, N. Walker, A. A. Saveliev, G. M. Smith, *Mixed Effects Models and Extensions in Ecology with R* (Springer, New York, 2009).
48. R Core Team, *R: A Language and Environment for Statistical Computing* (R Foundation for Statistical Computing, Vienna, 2016).